

## Aberystwyth University

### *Temperate airborne grass pollen defined by spatio-temporal shifts in community composition*

Brennan, Georgina L.; Potter, Caitlin; De Vere, Natasha; Griffith, Gareth; Skjøth, Carsten A.; Osborne, Nicholas J.; Wheeler, Benedict W.; McInnes, Rachel N.; Clewlow, Yolanda; Barber, Adam; Hanlon, Helen M.; Hegarty, Matthew; Jones, Laura; Kurganskiy, Alexander; Rowney, Francis M.; Armitage, Charlotte; Adams-Groom, Beverley; Ford, Col R.; Petch, Geoff M.; Consortium, PollerGEN

*Published in:*

Nature Ecology and Evolution

*DOI:*

[10.1038/s41559-019-0849-7](https://doi.org/10.1038/s41559-019-0849-7)

*Publication date:*

2019

*Citation for published version (APA):*

Brennan, G. L., Potter, C., De Vere, N., Griffith, G., Skjøth, C. A., Osborne, N. J., Wheeler, B. W., McInnes, R. N., Clewlow, Y., Barber, A., Hanlon, H. M., Hegarty, M., Jones, L., Kurganskiy, A., Rowney, F. M., Armitage, C., Adams-Groom, B., Ford, C. R., Petch, G. M., ... Creer, S. (2019). Temperate airborne grass pollen defined by spatio-temporal shifts in community composition. *Nature Ecology and Evolution*, 3(N/A), 750-754. <https://doi.org/10.1038/s41559-019-0849-7>

#### **Document License** CC BY-NC

#### **General rights**

Copyright and moral rights for the publications made accessible in the Aberystwyth Research Portal (the Institutional Repository) are retained by the authors and/or other copyright owners and it is a condition of accessing publications that users recognise and abide by the legal requirements associated with these rights.

- Users may download and print one copy of any publication from the Aberystwyth Research Portal for the purpose of private study or research.
- You may not further distribute the material or use it for any profit-making activity or commercial gain
- You may freely distribute the URL identifying the publication in the Aberystwyth Research Portal

#### **Take down policy**

If you believe that this document breaches copyright please contact us providing details, and we will remove access to the work immediately and investigate your claim.

tel: +44 1970 62 2400  
email: [is@aber.ac.uk](mailto:is@aber.ac.uk)

1   **Title: Temperate airborne grass pollen defined by spatio-temporal shifts in community**  
2   **composition**

3   **Authors:** Georgina L. Brennan<sup>1\*</sup>, Caitlin Potter<sup>2</sup>, Natasha de Vere<sup>2,3</sup>, Gareth W. Griffith<sup>2</sup>,  
4   Carsten A. Skjøth<sup>4</sup>, Nicholas J. Osborne<sup>5,6,7</sup>, Benedict W. Wheeler<sup>5</sup>, Rachel N. McInnes<sup>8</sup>,  
5   Yolanda Clewlow<sup>8</sup>, Adam Barber<sup>8</sup>, Helen M. Hanlon<sup>8</sup>, Matthew Hegarty<sup>2</sup>, Laura  
6   Jones<sup>1,3</sup>, Alexander Kurganskiy<sup>4</sup>, Francis M. Rowney<sup>5</sup>, Charlotte Armitage<sup>9</sup>, Beverley Adams-  
7   Groom<sup>4</sup>, Col R. Ford<sup>3</sup>, Geoff M. Petch<sup>4</sup>, The PollerGEN Consortium, and Simon Creer<sup>1\*</sup>.

8

9   **Consortium** Angela Elliot<sup>9</sup>, Carl A. Frisk<sup>4</sup>, Roy Neilson<sup>10</sup>, Stephen Potter<sup>11</sup>, Abdullah M. Rafiq<sup>1</sup>,  
10   David, B. Roy<sup>12</sup>, Katherine Selby<sup>13</sup>, Natascha Steinberg<sup>9</sup>

11

12   **Affiliations:**

13   <sup>1</sup>Molecular Ecology and Fisheries Genetics Laboratory, School of Natural Sciences, Bangor  
14   University, Bangor, Gwynedd, Wales, UK

15   <sup>2</sup>IBERS, Aberystwyth University, Aberystwyth, Ceredigion, Wales, UK

16   <sup>3</sup>National Botanic Garden of Wales, Llanarthne, Carmarthenshire, Wales, UK

17   <sup>4</sup>University of Worcester, Worcester, Worcestershire, UK

18   <sup>5</sup>University of Exeter, Truro, Cornwall, UK

19   <sup>6</sup>University of New South Wales, Sydney, New South Wales, Australia

20   <sup>7</sup>School of Public Health, Faculty of Medicine, The University of Queensland, Herston,  
21   Brisbane, Australia

22 <sup>8</sup>Met Office, Exeter, Devon, UK

23 <sup>9</sup>The Woodland Trust, Kempton Way, Grantham, Lincolnshire, UK.

24 <sup>10</sup>College of Life and Environmental Science, University of Exeter, Exeter, UK

25 <sup>11</sup>Ecological Sciences, The James Hutton Institute, Dundee, Scotland, UK

26 <sup>12</sup>The David Hide Asthma & Allergy Research Centre, St Mary's Hospital, Newport, Isle of  
27 Wight, UK

28 <sup>13</sup>Centre for Ecology & Hydrology, Crowmarsh Gifford, Wallingford, Oxfordshire, UK.

29 <sup>14</sup>Environment Department, University of York, Heslington, York, UK

30

31 **Corresponding authors:** Simon Creer ([s.creer@bangor.ac.uk](mailto:s.creer@bangor.ac.uk)) and Georgina L. Brennan

32 ([g.l.b.doonan@gmail.com](mailto:g.l.b.doonan@gmail.com))

33

34 Grass pollen is the world's most harmful outdoor aeroallergen, yet it is unknown how  
35 airborne pollen assemblages change in time and space. Human sensitivity varies between  
36 different species of grass that flower at different times, but it is not known if temporal  
37 turnover in species composition match terrestrial flowering or if species richness steadily  
38 accumulates over the grass pollen season. Here, using targeted, high-throughput sequencing,  
39 we demonstrate that all grass genera display discrete, temporally restricted peaks of  
40 incidence which varied with latitude and longitude throughout Great Britain, revealing that  
41 the taxonomic composition of grass pollen exposure changes substantially across the grass  
42 pollen season.

43  
44 Allergens carried in airborne pollen are associated with both asthma<sup>1</sup> and allergic rhinitis (hay  
45 fever), negatively affecting 400 million people worldwide<sup>2</sup>. Pollen from the grass family  
46 (Poaceae) constitutes the most significant outdoor aeroallergen<sup>3,4</sup>, and more people are  
47 sensitised to grass pollen than to any other pollen type<sup>5</sup>. However, despite the harmful  
48 impact of grass pollen on human health, especially as reported in developed nations, current  
49 studies and forecasts categorize grass pollen at the family level (Poaceae)<sup>6,7</sup> due to difficulties  
50 in differentiating species based on morphology<sup>8</sup>. Furthermore, while different species of  
51 temperate grass flower at different timepoints<sup>9,10</sup>, it is unknown if the disparate phenology of  
52 local grass taxa at ground level are useful for making predictions on the seasonal variation in  
53 airborne pollen. Airborne pollen is highly mobile<sup>11,12</sup> and pollen concentrations often do not  
54 directly correlate to local flowering times<sup>11</sup>. Persistence and mobility of grass pollen could  
55 result in steadily increasing species richness of airborne pollen over the grass pollen season.  
56 Conversely, if grass pollen does not persist for an extended time in the air, pollen  
57 assemblages should reflect temporal turnover in species composition over the summer

months. Understanding the taxon-specific phenology of airborne pollen would fill a significant knowledge gap in our understanding of allergen triggers, with associated benefits to healthcare providers, pharmaceutical industries and the public.

Many species within the subfamilies Pooideae, Chloridoideae, and Panicoideae release allergenic pollen into the atmosphere<sup>5</sup>, including *Phleum* spp. (e.g. Timothy grasses), *Dactylis* spp. (Cocksfoot/Orchard grasses), *Lolium* spp. (Ryegrasses), *Festuca* spp. (Fescues), *Poa* spp. (Meadow-grasses and Bluegrasses), and *Anthoxanthum* spp. (Vernal grasses). Furthermore, some grass taxa, notably diverse cultivars and hybrids of *Lolium* spp., are widely sown in agricultural grasslands and are likely to contribute disproportionately to airborne pollen. However, it is unknown whether particular grass species, or varieties/cultivars within species, contribute more to the prevalence of allergic symptoms and related diseases than others<sup>13</sup>.

Whilst some grass species have been identified as more allergenic than others *in vitro* (triggering higher levels of Immunoglobulin E (IgE) antibody production), there is a high degree of cross-reactivity between grass species<sup>14–16</sup>. In addition, the allergen profiles (the characterisation of the different allergens common to different grass), the degree of sensitisation differ between grass species<sup>14,17</sup>, and the overall allergenicity of grass pollen in the air varies across seasons<sup>18</sup>. Family-level estimates of grass pollen concentrations cannot therefore be considered a reliable proxy for either the concentration of pollen-derived aeroallergens or pollen-induced public health outcomes.

The identification of biodiversity via the high-throughput analysis of taxonomic marker genes (popularly termed metabarcoding) provides an emerging solution to semi-quantitatively identify complex mixtures of airborne pollen grains<sup>19–22</sup>. Further, recent global DNA

82 barcoding initiatives and co-ordinated regional efforts have now resulted in near complete  
83 genetic databases of national native plants, including grass species. In Great Britain, the vast  
84 majority of angiosperms are included in mature DNA barcoding databases for multiple  
85 markers<sup>23</sup>, meaning that we are now in a position to investigate the aerial composition of  
86 pollen over the grass pollen season, at a national scale.

87

88 Here, using two complementary DNA barcode marker genes (*rbcL* and ITS2), we characterise  
89 the spatial and temporal distribution of airborne grass pollen throughout the temperate  
90 summer grass pollen season (May-August) across the latitudinal and longitudinal range of  
91 Great Britain (Fig. 1). We hypothesise that the composition of airborne grass pollen, from  
92 different grass taxa will be (i) broadly homogenous across the grass pollen season, regardless  
93 of terrestrial Poaceae phenology and (ii) homogenous across Great Britain due to the  
94 potential for long distance transport of windborne pollen grains.

95

96 Airborne grass pollen from each genus occupied distinct temporal windows across the grass  
97 pollen season in 2016 (May to August), thereby rejecting our hypothesis (i) (Fig. 2,  
98 Supplementary Figure 1). Time, measured as number of days after the first sample was  
99 collected, is a good predictor of airborne grass pollen taxon composition using both markers  
100 (Fig. 2, Supplementary Figure 1; *ITS2*,  $LR_{1,74} = 128.8$ ,  $P = 0.001$ ; *rbcL*,  $LR_{1,71} = 46.71$ ,  $P =$   
101  $0.001$ ). Community-level ordination reveals that the airborne grass pollen community as a  
102 whole changed across the grass pollen season (Supplementary Figure 2, Supplementary  
103 Figure 3), with similar overarching trends observed for the most abundant airborne pollen  
104 families including, Poaceae, Pinaceae and Urticaceae (Supplementary Figure 4). In addition,  
105 observations of first flowering dates from a citizen science project (UKPN;

[www.naturescalendar.org.uk](http://www.naturescalendar.org.uk)) and metabarcoding data show similar sequences of seasonal progression with a lag time similar to that found in observational studies<sup>11</sup> (See supplementary text; Supplementary Figure 5), suggesting that there is a link between local phenology of Poaceae and composition of airborne grass pollen.

Focusing on the more taxonomically specific ITS2 marker dataset, *Alopecurus* and *Holcus* typically dominated the early grass pollen season (Fig. 2), which coincides with typical peaks in allergic rhinitis<sup>24</sup>, but further research will be required to confirm this association. *Lolium* featured prominently for the majority of the later grass season. The popularity of *Lolium* species as a forage crop means that it is widely sown in agricultural grasslands<sup>25</sup>, although the majority of agricultural grasslands are managed by grazing silage-cutting or mowing which prevents the growth of flowering heads<sup>25</sup>. The length of time over which *Lolium* pollen dominated may be because many varieties have been bred with the potential to mature at different times throughout the year<sup>26</sup>, although it should be noted that *Lolium* species frequently hybridise with each other and therefore it is difficult to distinguish these genera using genetic material alone. Additionally, while there is some evidence that some species of grass appear to be more allergenic than others<sup>18</sup>, it is unknown how much they may differ within a species (i.e. at the cultivar/hybrid level)<sup>16</sup>. Although *Lolium* was the dominant species in airborne grass pollen from July to the end of the sampling period, the total grass pollen concentration declined in August, indicating that the absolute number of *Lolium* pollen grains at this time is low (Fig. 1, Supplementary Figure 6).

The top five genera contributing to airborne pollen, indicated by the relative abundance of taxonomy marker genes, were *Alopecurus*, *Festuca*, *Holcus*, *Lolium* and *Poa* (Fig. 2;

Supplementary Figure 6). Each of these genera is widespread in the UK, although long-distance pollen transport means they may also originate further afield<sup>27</sup>. These dominant genera have all been shown to provoke IgE-mediated responses in grass-sensitised patients<sup>14</sup>, providing candidate species for links with hay fever and asthma exacerbation. Conversely, less prevalent species in the dataset could contribute disproportionately to the allergenic load. Species such as *Phleum pratense* have been identified to be a major allergen<sup>5,28</sup>. However, we found that *Phleum* made up a very small proportion of metabarcoding reads (Supplementary Figure 1), corresponding with the results of an earlier phenological study<sup>9</sup>. Most genera, such as *Phleum*, *Anthoxanthum* and *Dactylis*, show distinct and narrow temporal incidence (Supplementary Figure 1), and could allow researchers to identify grass species associated with allergenic windows with greater accuracy.

Changes in species composition over time were localised. We found that peaks in abundance of airborne pollen occurred at different times at each location during the summer (Fig. 2, Supplementary Figure 1). For example, the relative abundance of airborne grass pollen from the genus *Poa* peaked in mid-June in Worcester and Bangor but 6-8 weeks later in Invergowrie (Fig. 2), probably due to latitudinal effects on flowering time<sup>7,27</sup>. This is supported by a significant interaction between latitude and time of year for both markers (Fig. 2, Supplementary Figure 1; *ITS2*,  $LR_{68,1} = 34.2$ ,  $P = 0.002$ ; *rbcL*,  $LR_{68,1} = 47.36$ ,  $P = 0.001$ ).

Differences in species composition of airborne grass pollen between the six sampling sites is supported by a significant effect of latitude (Fig. 2, Supplementary Figure 1; *ITS2*,  $LR_{1,73} = 73.2$ ,  $P = 0.001$ ; *rbcL*,  $LR_{1,70} = 26.4$ ,  $P = 0.025$ ) and longitude (Fig. 2, Supplementary Figure 1; *ITS2*,  $LR_{1,69} = 33$ ,  $P = 0.003$ ; *rbcL*,  $LR_{1,69} = 27.10$ ,  $P = 0.010$ ), that are proxies for a broad range of environmental variables. These results do not support our hypothesis (ii) that the



composition of airborne grass pollen will be homogenous across the UK, and instead suggest that taxon-specific effects of regional geography, climate and environmental conditions underpin distributions which have been demonstrated for Poaceae pollen as a whole<sup>7</sup>. Further investigations into the mechanisms of pollen production and transport, interacting with a range of climatic, seasonal and meteorological effects will therefore provide valuable future research foci to elucidate our mechanistic knowledge of the deposition of grass pollen in time and space.

Enabled by contemporary molecular biodiversity assessment and mature, curated DNA barcoding databases, here we provide a comprehensive taxonomic overview of airborne grass pollen distribution, throughout an entire grass pollen season and across large geographic scales. The grass pollen season is defined by discrete temporal windows of different grass species, with some species displaying geographical variation. Temporal pollen distributions in metabarcoding data follow observed flowering times. The data provide an important step towards developing genera-, and in certain cases, species-level grass pollen forecasting. Additionally, the research presented here leads the way for future studies facilitating understanding of the relationships between grass pollen and disease, which have significant global public health relevance and socioeconomic importance.

**Figure 1 Location of pollen collection and temporal Poaceae concentrations and composition.**

Map showing location of the six sampling sites and daily Poaceae pollen concentrations (grains/m<sup>3</sup>) throughout the grass pollen season (May to August, 2016). Yellow filled circles indicate dates when pollen was collected for both observational concentrations and molecular analysis, blue circles indicate days when pollen was collected for observational

concentrations only and green circles indicate when pollen was collected for molecular analysis only at the Bangor site. Note that Bangor is not part of the UK pollen monitoring network and observational concentrations were only performed alongside pollen collections for molecular analysis between 24<sup>th</sup> June to 28<sup>th</sup> August 2016. Contains OS data Crown copyright and database right (2018). Image Crown Copyright, 2018, The Met Office.

**Figure 2 Abundance of the most common airborne grass pollen taxa throughout the grass pollen season.** The five most abundant grass taxa (expressed as proportion of total reads), depicted alongside the total proportion of reads assigned to family Poaceae. Due to errors in sampling equipment, only 4 alternate weeks (out of a possible 7 alternate weeks) of samples were collected at the York sampling site. Markers used to identify grass pollen are stated in the top panel label. Sampling sites are indicated in the right panel label abbreviated as follows: BNG = Bangor; EXE = Exeter; ING = Invergowrie; IOW = Isle of Wight; WOR = Worcester; YORK = York. A map of sampling locations and daily Poaceae pollen concentrations can be found in Figure 1.

## **Methods**

### **Sampling and Experimental Design**

We collected aerial samples from six sites across Great Britain (Supplementary Table 3; Fig. 1) using Burkard Automatic Multi-Vial Cyclone Samplers (V2; Burkard Manufacturing Co. Ltd. Rickmansworth, UK) designed to simplify collection of pollen and spores by sampling directly into a microcentrifuge tube (e.g.<sup>29</sup>). The volumetric aerial sampler uses a turbine to draw in air (16.5 litres/min) and aerial particles, using mini-cyclone technology. The aerial particles

are collected into 1.5 ml sterile microcentrifuge tubes located on a carousel, where the carousel is programmed to sample into a new tube every 24 h, thereby providing daily samples of airborne pollen (Supplementary Figure 7). Sample tubes were sent to Bangor, and stored at -20°C before processing. Each sampling unit was mounted alongside a seven-day volumetric trap of the Hirst design (1952) belonging to the Met Office UK Pollen Monitoring Network, which provided daily pollen concentrations (Fig. 1; map produced using ArcGIS). In the seven-day volumetric trap, a turbine draws air in (10 litres/min) and particles are impacted upon an adhesive coated tape carried on a clockwork-driven drum. The tape is cut into 24 h sections, and mounted on glass slides using a gelatine/glycerol mountant containing basic fuchsin to stain the pollen grains. Pollen are identified and counted under a microscope and converted to volumetric concentrations<sup>7</sup>. Although the high cost of the pollen samplers preclude routine replicate sampling, our methodologies mirror methodologies that have been used for several decades in the UK network<sup>30,31</sup> and are in agreement with recommended terminology described by Galan et al (2017)<sup>32</sup>. All pollen samplers were sited in elevated positions on flat-roofed buildings between 4 to 6 floors in height in order to sample from a mixed air flow. Fins on the samplers (both Burkard Multi-Vial Cyclone and Hirst type seven-day volumetric samplers) direct the cyclone inlet port into the wind. Bangor was the only sampling site which was not part of the pollen monitoring network, but we deployed the same methodology at the Bangor site (which began on 24<sup>th</sup> June 2016 Fig. 1).

Sampling began in late May 2016 and during alternate weeks, aerial samples were collected for seven days for a total of seven weeks between 25<sup>th</sup> May and 28<sup>th</sup> August. Exact sampling dates varied slightly between sites and a total of 279 aerial samples were collected (Supplementary Table 4).

## DNA Extraction, PCR and Sequencing

From the 279 daily aerial samples, 231 were selected for downstream molecular analysis, as described below. Within each sampling week, two series of three consecutive days were pooled. Pooled samples were selected based on grass pollen concentrations based on microscopy. The final, unselected day was not used in downstream molecular analysis. In total, seventy-seven pools of DNA were created. In one instance, three consecutive days of pollen samples were unavailable (Invergowrie, week 2, pool 2) due to trap errors. For this sample, the next sampling day was selected for pooling (Supplementary Table 4). DNA was extracted from daily samples using a DNeasy Plant Mini kits (Qiagen, Valencia, CA, USA), with some modifications to the standard protocol as described by Hawkins *et al.*<sup>33</sup>. DNA from daily samples was pooled and eluted into 60 µl of elution buffer at the binding stage of the DNeasy Plant Mini kit.

Illumina MiSeq paired end indexed amplicon libraries were prepared following a two-step protocol. Two marker genes were amplified with universal primer pairs *rbcLaf* and *rbcLr506*<sup>23,34</sup>, and ITS2 and ITS3<sup>18</sup> (Supplementary Table 5). A 5' universal tail was added to the forward and reverse primers and a 6N sequence was added between the forward universal tail and the template-specific primer, which is known to improve clustering and cluster detection on MiSeq sequencing platforms<sup>35</sup> (Integrated DNA Technologies, Coralville, USA). Round 1 PCR was carried out in a final volume of 25 µL, including forward and reverse primers (0.2 µM), 1X Q5 HS High-Fidelity Master Mix (New England Biolabs) and 1 µL of template DNA. Thermal cycling conditions were an initial denaturation step at 98 °C for 30s; 35 cycles of 98 °C for 10s, 50 °C for 30s, 72 °C for 30s; and a final annealing step of 72 °C for 5

minutes. Products from the first PCR were purified using Agencourt AMPure XP beads (Beckman Coulter) with a 1:0.6 ratio of product to AMPure XP beads.

The second round PCR added the unique identical i5 and i7 indexes and the P5 and P7 Illumina adaptors, along with universal tails complementary to the universal tails used in round 1 PCR (Supplementary Table 4, Supplementary Table 5) (Ultramer, by IDT, Integrated DNA Technologies). Round 2 PCR was carried out in a final volume of 25 µL, including forward and reverse index primers (0.2 µM), 1X Q5 HS High-Fidelity Master Mix (New England Biolabs) and 5 µL of purified PCR product. Thermal cycling conditions were: 98 °C for 3 min; 98 °C for 30 s, 55 °C for 30 s, 72 °C for 30 s (10 cycles); 72 °C for 5 min, 4 °C for 10 min. Both PCRs were run in triplicate. The same set of unique indices were added to the triplicates which were then pooled following visual inspection on an agarose gel (1.5%) to ensure that indices were added successfully. Pooled metabarcoding libraries were cleaned a second time using Agencourt AMPure magnetic bead purification, run on an agarose gel (1.5%) and quantified using the Qubit high sensitivity kit (Thermo Fisher Scientific, Massachusetts, USA). Positive and negative controls were amplified in triplicate with both primer pairs and sequenced alongside airborne plant community DNA samples using the MiSeq. Sequence data, including metadata, are available at the Sequence Read Archive (SRA) using the project accession number SUB4136142.

## **Bioinformatic Analysis**

Initial sequence processing was carried out following a modified version of the workflow described by de Vere *et al.*<sup>36</sup>. Briefly, raw sequences were trimmed using Trimmomatic v0.33<sup>37</sup> to remove short reads (<200bp), adaptors and low quality regions. Reads were

merged using FLASH v 1.2.11<sup>36,38</sup>, and merged reads shorter than 450bp were excluded. Identical reads were merged using fastx-toolkit (v0.0.14), and reads were split into ITS2 and *rbcL* based on primer sequences.

To prevent spurious BLAST hits, custom reference databases containing *rbcL* and ITS2 sequences from UK plant species were generated. While all native species of the UK have been DNA barcoded<sup>23</sup>, a list of all species found in the UK was generated in order to gain coverage of non-native species. A list of UK plant species was generated by combining lists of native and alien species<sup>39</sup> with a list of cultivated plants obtained from Botanic Gardens Conservation International (BGCI) which represented horticultural species. All available *rbcL* and ITS2 records were downloaded from NCBI GenBank, and sequences belonging to UK species were extracted using the script '[creatingselectedfastadatabase.py](#)', archived on GitHub.

Metabarcoding data was searched against the relevant sequence database using blastn<sup>40</sup>, via the script 'blast\_with\_ncbi.py'. The top twenty blast hits (identified using the highest bit-score) were tabulated ('blast\_summary.py'), then manually filtered to limit results to species currently present in Great Britain. Reads occurring fewer than four times were excluded from further analysis. All scripts used are archived on GitHub: <https://doi.org/10.5281/zenodo.1305767>.

## Statistical Analysis

To understand how the grass pollen composition changed with space and time, the effect of time (measured as the number of days after the first sampling date), latitude and longitude

of sampling location were included in a two-tailed generalized linear model using the 'manyglm' function in the package 'mvabund'<sup>41</sup>. The proportion of sequences was set as the response variable; proportion data was used as this has been shown to be an effective way of controlling for differences in read numbers<sup>42</sup>. The effect of time, latitude, longitude and the interaction between time and latitude were included as explanatory variables in the models to test hypotheses (i and ii). The effect of longitude is also consistent when York, the most easterly sampling site, with missing data from mid-July until the end of the sampling period, is removed from the analysis (Supplementary Table 6).

The data best fit a negative binomial distribution, most likely due to the large number of zeros (zeros indicate that a grass genus is absent from a sample), resulting in a strong mean-variance relationship in the data (Supplementary Figure 8). The proportion of sequences was scaled by 1000 and values were converted to integers so that a generalized linear model with a negative binomial distribution could be used. Overfitting of the models was tested using 'dropterm' in R, and based on the lowest Akaike Information Criterion (AIC) score, no terms were removed from the models. In addition, the appropriateness of the models was checked by visual inspection of the residuals against predicted values from the models (Supplementary Figure 9).

In order to compare the metabarcoding data with flowering time data, we used phenological records of first flowering collected in 2016 by citizen scientists from the UK's Nature's Calendar ([www.naturescalendar.org.uk](http://www.naturescalendar.org.uk)). First flowering time was compared to genus-level ITS2 metabarcoding data for three species: *Alopecurus pratensis*, *Dactylis glomerata* and *Holcus lanatus*. As grass pollen could only be reliably identified to genus level in the

metabarcoding data, the taxa compared may not have been exactly equivalent since both *Alopecurus* and *Holcus* contain other widespread species within the UK. However, *Alopecurus pratensis* and *Holcus lanatus* are the most abundant species within their respective genera. The comparison was only carried out for ITS2 data because two of the three genera were not identified by the *rbcl* marker.

NMDS ordination was carried out using package 'VEGAN' in R<sup>43</sup>, based on the proportion of total high-quality reads contributed by each grass genus, using Bray-Curtis dissimilarity (Supplementary Figure 2 and Supplementary Figure 3). Ordination is used to reduce multivariate datasets (e.g. abundances of many species) into fewer variables that reflect overall similarities between samples. A linear model was carried out using the 'lm' function within the 'stats' package in R, in order to investigate the relationship between the number of reads obtained for each genus using the *rbcl* and ITS2 marker.

**Data and materials availability:** All sequence data are available at the Sequence Read Archive (SRA) using the project accession number SUB4136142. Archived sequence data was used to generate Figure 2 Supplementary Figures 1-S6 and 8-S10). First flowering data used in Supplementary Figure 5 was obtained from Nature's Calendar, Woodland Trust and is available upon request. The sequence analysis pipeline is available at <https://github.com/colford/nbgw-plant-illumina-pipeline>.

## References

1. Blackley, C.H. (London: Bailliere: London, 1873).
2. Marks, G., Pearce, N., Strachan, D. & Asher, I. *The Global Asthma Report 2014* 16–21



- 346 (2014).
- 347 3. Bauchau, V. *The European Respiratory Journal* **24**, 758–764 (2004).
- 348 4. Bousquet, P.-J. et al. *Allergy* **62**, 301–309 (2007).
- 349 5. García-Mozo, H. *Allergy* **72**, 1849–1858 (2017).
- 350 6. Emberlin, J. et al. *Aerobiologia* **16**, 373–379 (2000).
- 351 7. Emberlin, J. et al. *Grana* **33**, 94–99 (1994).
- 352 8. Mander, L., Li, M., Mio, W., Fowlkes, C.C. & Punyasena, S.W. *Proceedings of The Royal*  
353 *Society B* **280**, 20131905 (2013).
- 354 9. Kmenta, M. et al. *World Allergy Organ. J.* **10**, 31 (2017).
- 355 10. Cope, C. & Gray, A. (Botanical Society of the British Isles: London, UK, 2009).
- 356 11. Estrella, N., Menzel, A., Krämer, U. & Behrendt, H. *International Journal of*  
357 *Biometeorology* **51**, 49–59 (2006).
- 358 12. Skjøth, C.A., Sommer, J., Stach, A., Smith, M. & Brandt, J. *Clinical & Experimental*  
359 *Allergy* **37**, 1204–1212 (2008).
- 360 13. McInnes, R.N. et al. *Science of the Total Environment* **599–600**, 483–499 (2017).
- 361 14. van Ree, R., van Leeuwen, W.A. & Aalberse, R.C. *Journal of Allergy and Clinical*  
362 *Immunology* **102**, 184–190 (1998).
- 363 15. Petersen, A. et al. *Journal of Allergy and Clinical Immunology* **107**, 856–862 (2001).
- 364 16. Jung, S. et al. *PLoS ONE* **13**, 1–12 (2018).
- 365 17. Moingeon, P., Peltre, G. & Bergmann, K.-C. *Clinical and Experimental Allergy Reviews*  
366 **8**, 12–14 (2008).
- 367 18. de Weger, L.A. et al. *Clinical and Translational Allergy* **1**, 18 (2011).
- 368 19. Kraaijeveld, K. et al. *Molecular Ecology Resources* **15**, 8–16 (2015).
- 369 20. Korpelainen, H. & Pietiläinen, M. *Nordic Journal of Botany* **35**, 602–608 (2017).
- 370 21. Deiner, K. et al. *Molecular Ecology* **26**, 5872–5895 (2017).
- 371 22. Creer, S. et al. *Methods in Ecology and Evolution* **7**, 1008–1018 (2016).
- 372 23. de Vere, N. et al. *PLoS ONE* **7**, 1–12 (2012).
- 373 24. England, P.H. (PHE Real-time Syndromic Surveillance Team: Birmingham, UK, 2016).

- 374 25. Frame, J. & Laidlaw, A.S. (The Crowood Press: Marlborough, Wiltshire, UK, 2011).
- 375 26. RGCL (British Grassland Society: Cheshire, UK, 2017).
- 376 27. Rousseau, D.-D. et al. *Geophysical Research Letters* **30**, (2003).
- 377 28. D'Amato, G. et al. *Allergy and Allergen Immunotherapy* **62**, 261–279 (2017).
- 378 29. West, J.S. & Kimber, R.B.E. *Annals of Applied Biology* **166**, 4–17 (2015).
- 379 30. Adams-Groom, B., Emberlin, J., Corden, J., Millington, W. & Mullins, J. *Aerobiologia*  
380 **18**, 117–123 (2002).
- 381 31. Skjøth, C.A., Baker, P., Sadyś, M. & Adams-Groom, B. *Urban Climate* **14**, 414–428  
382 (2015).
- 383 32. Galán, C. et al. *Aerobiologia* **33**, 293–295 (2017).
- 384 33. Hawkins, J. et al. *PLoS ONE* **10**, 1–20 (2015).
- 385 34. Kress, W.J. & Erickson, D.L. *PLoS One* **2**, e508 (2007).
- 386 35. Miya, M. et al. *Royal Society open science* **2**, 150088 (2015).
- 387 36. de Vere, N. et al. *Scientific Reports* **7**, 42838 (2017).
- 388 37. Bolger, A.M., Lohse, M. & Usadel, B. *Bioinformatics* **30**, 2114–2120 (2014).
- 389 38. Magoc, T. & Salzberg, S.L. *Bioinformatics* **27**, 2957–2963 (2011).
- 390 39. Stace, C. (Cambridge University Press: Cambridge, UK, 2010).
- 391 40. Camacho, C. et al. *BMC Bioinformatics* **10**, 421 (2008).
- 392 41. Wang, Y., Naumann, U., Wright, S.T. & Warton, D.I. *Methods in Ecology and Evolution*  
393 **3**, 471–474 (2012).
- 394 42. McMurdie, P.J. & Holmes, S. *PLoS Computational Biology* **10**, e1003531 (2014).
- 395 43. Dixon, P. *Journal of Vegetation Science* **14**, 927–930 (2003).

396

## 397 Acknowledgements

398

399 We thank John Kenny, Pia Koldkjær, Richard Gregory, and Anita Lucaci of the Liverpool  
400 Centre for Genomic Research for sequencing support. We acknowledge the computational  
401 services & support of the Supercomputing Wales project, which is part-funded by the

European Regional Development Fund (ERDF) via Welsh Government. We thank the Botanic Gardens Conservation International (BGCI) for access to the list of plant collections in the National Gardens in the UK and Ireland. We thank the Met Office network for providing additional observational grass pollen count data and Jonathan Winn, UK Met Office for ArcGIS assistance on Figure 1. We are grateful to the Woodland Trust and Centre for Ecology & Hydrology for supplying the UK Phenology Network data and to the citizen scientists who have contributed to the latter scheme. Final thanks to Wendy Grail and technical support staff at Bangor University.

**Author Contributions:** S.C., N.dV., G.W.G., R.N.M., N.J.O., C.A.S., Y.C., B.W.W. and G.L.B. conceived and designed the study; B.A-G., G.L.B., G.P., A.E., R.N., S.P., K.S., and N.S. collected samples and counted pollen; G.L.B. performed laboratory work, supported by S.C.; N.dV., C.P., C.R.F., L.J., S.C. contributed methods, and C.A. and D.B.R. contributed materials. C.P. and G.L.B. analysed the data and G.L.B., C.P. and S.C. produced the first draft of the manuscript. All authors contributed substantially to the final submitted manuscript.

**Funding:** This work was supported by the Natural Environment Research Council (<https://nerc.ukri.org/>), awarded to SC (NE/N003756/1), CS (NE/N002431/1), NO (NE/N002105/1) and GWG/NdV/MH (NE/N001710/1). IBERS Aberystwyth receives strategic funding from the BBSRC. The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

**Competing interests:** The authors are not aware of any competing interests.





